

# Introduction to Bioinformatics

Jianlin Cheng, PhD

Department of Computer Science  
Informatics Institute



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# Topics

- Introduction
- Biological Sequence Alignment and Database Search
- Analysis of gene expression data

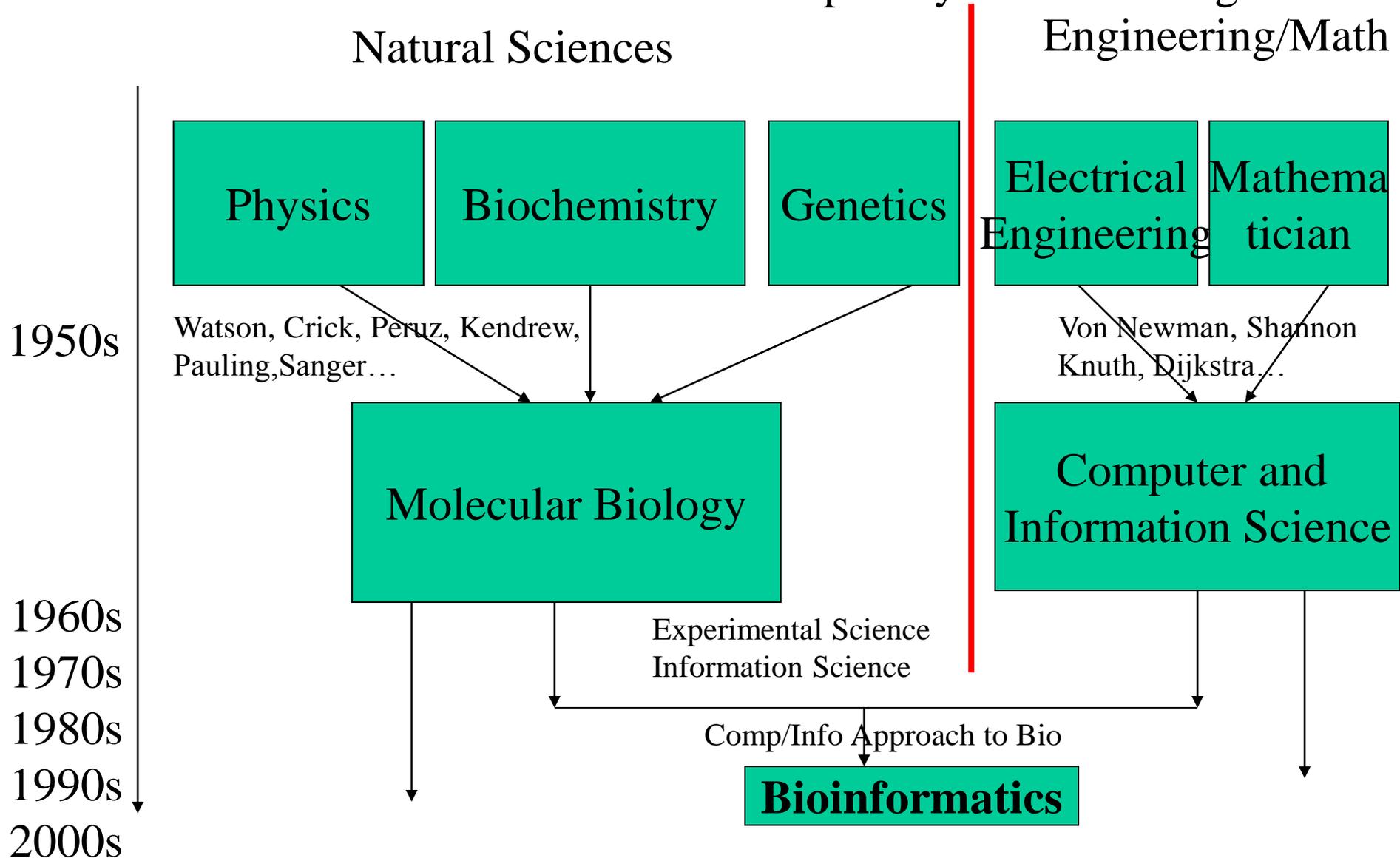
# What's Bioinformatics?

An interdisciplinary science of developing and applying computational techniques to address problems in molecular biology

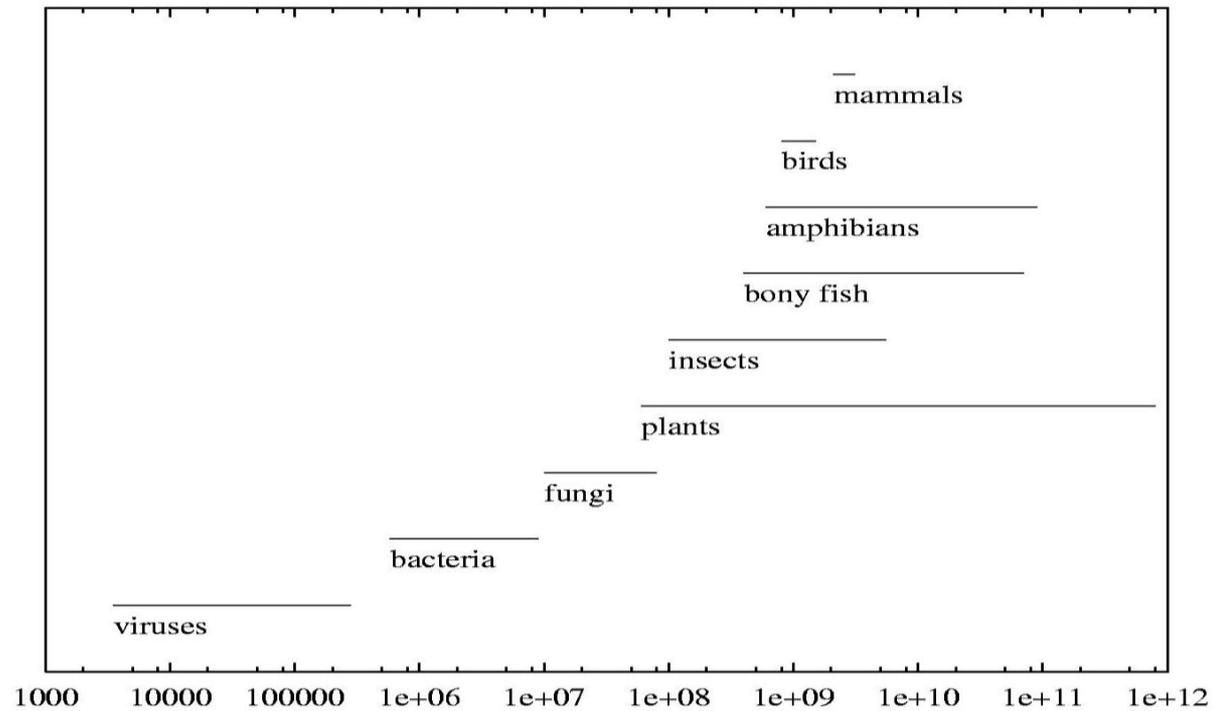
- Develop bioinformatics algorithms and tools
- Apply bioinformatics tools to address biological problems

# History of Bioinformatics

How does a new interdisciplinary science emerge?



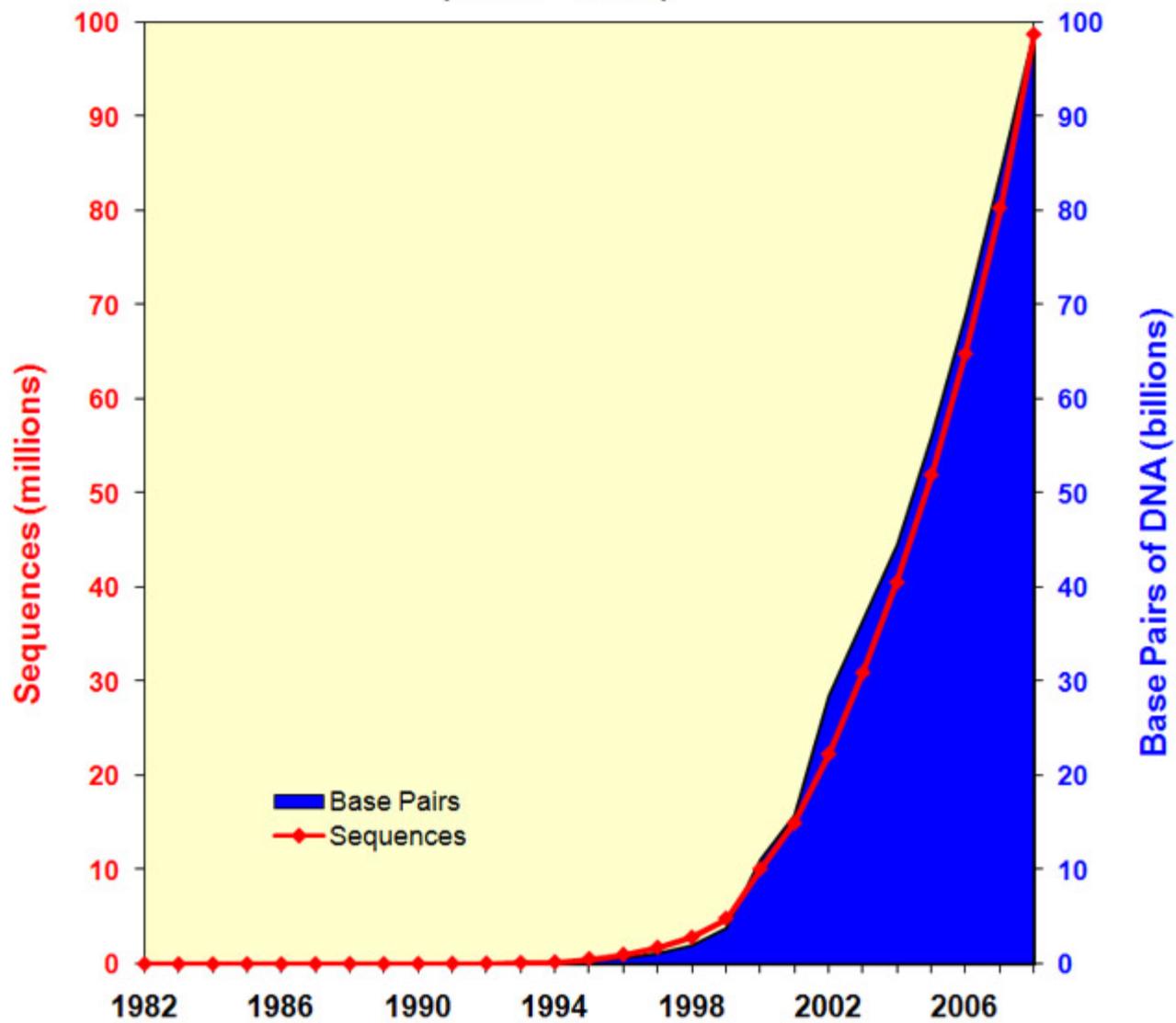
# Genome Sequencing



# High-Throughput Sequencing

- Transcriptome (EST, RNA-Seq, Chip-Seq)
- Proteomics (Mass Spectrometry)
- Metabolomics

# Growth of GenBank (1982 - 2008)



# What can we do with these huge amount of data?



**Find buried treasure - Doug Brutlag, 1999.**

# Typical Bioinformatics Problems

- What family does this gene / protein belong to?
- Are there other known homologous proteins?
- What is the function and structure of this protein?
- What biological pathway does this protein participate in?
- Is a mutation on a gene / protein related to a phenotype or disease?
- Is a gene differentially expressed in a biological condition?

# Fundamental Problems: Sequence Comparison

- Why do we compare sequences?
- What's similarity between two sequences?
- How to compare sequences?
- Is similarity significant?

# Importance of Similarity Comparison

- Identify evolutionary relationship between genes and proteins
- Similar genes/proteins have similar function
- Similar proteins have similar structures



# Three Main Issues

1. Definition of alignment score
2. Algorithms of finding the optimal alignment
3. Evaluation of significance of alignment score

# A simple scoring scheme

- Score of character pair:  $S(\text{match})=1$ ,  $S(\text{not\_match}) = -1$ ,  $S(\text{gap-char}) = -1$
- Score of an alignment =  $\sum_1^n S_i$

ITAKPAKTPTSPEQAIGLSVTFLSFLLPAGWVLYHL

ITAKPQWLKSTE-----SVTFLSFLLPQTQGLYHL

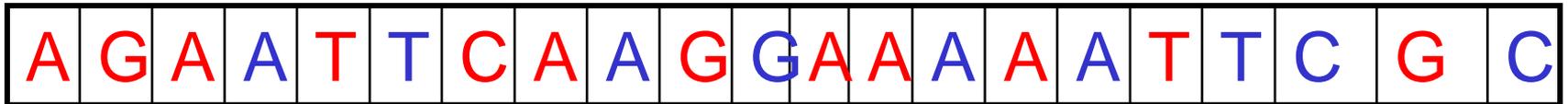
$$5 - 7 - 7 + 10 - 4 + 4 = 1$$

# Optimization

- How can we find the best alignment to maximize alignment score?
- How many possible alignments exist for two sequences with length  $m$  and  $n$ ?

# Total Number of Possible Alignments

AGATCAGAAAT-G  
--AT-AG-AATCC



**m + n**

# Total Number of Alignments

Select  $m$  positions out of  $m+n$  possible positions:

$$\binom{m+n}{m} = \frac{(m+n)!}{m!n!}$$

**Exponential!**

If  $m = 300$ ,  $n = 300$ , total =  $10^{37}$

# Divide and Conquer

Goal: align prefix P[1..i] and prefix Q[1..j]

i

Seq P: **AGATCAGAAATGG**

Seq Q: **ATAGAATCC**

j

Three possibilities assuming we know the optimal alignment of smaller prefixes:

## Case 1

Use alignment of P[1..i-1] and Q[1..j-1], pair P[i] and Q[j]

i

|         |
|---------|
| AGATCAG |
| --AT-AG |

j

## Case 2

Use alignment of P[1..i] and Q[1..j-1], pair Q[j] with gap

i

|          |
|----------|
| AGATCAG- |
| --AT-A-G |

j

## Case 3

Use alignment of P[1..i-1] and Q[1..j], pair P[i] with gap.

i

|          |
|----------|
| AGATCA-G |
| --AT-AG- |

j

# Needleman and Wunsch Algorithm

- Given sequences P and Q, we use a matrix M to record the optimal alignment scores of all prefixes of P and Q.  $M[i,j]$  is the best alignment score for the prefixes  $P[1..i]$  and  $Q[1..j]$ .
- $M[i,j] =$   
    **max** [  
         $M[i-1,j-1] + S(P[i],Q[j])$ ,  
         $M[i,j-1] + S(-, Q[j])$   
         $M[i-1,j] + S(P[i], -)$   
    ]

## Dynamic Programming

# Dynamic Programming Algorithm

## **Three-Step Algorithm:**

- Initialization
- Matrix fill (scoring)
- Trace back (alignment)

# 1. Initialization of Matrix M

|   |   |     |    |    |    |    |    |    |    |
|---|---|-----|----|----|----|----|----|----|----|
|   |   |     |    | j  |    |    |    |    |    |
|   | – | A   | T  | A  | G  | A  | A  | T  |    |
| i | – | 0   | -1 | -2 | -3 | -4 | -5 | -6 | -7 |
|   | A | -1  |    |    |    |    |    |    |    |
|   | G | -2  |    |    |    |    |    |    |    |
|   | A | -3  |    |    |    |    |    |    |    |
|   | T | -4  |    |    |    |    |    |    |    |
|   | C | -5  |    |    |    |    |    |    |    |
|   | A | -6  |    |    |    |    |    |    |    |
|   | G | -7  |    |    |    |    |    |    |    |
|   | A | -8  |    |    |    |    |    |    |    |
|   | A | -9  |    |    |    |    |    |    |    |
|   | A | -10 |    |    |    |    |    |    |    |
|   | T | -11 |    |    |    |    |    |    |    |
|   | G | -12 |    |    |    |    |    |    |    |

## 2. Fill Matrix

|   | -   | A   | T  | A  | G  | A  | A  | T  |
|---|-----|-----|----|----|----|----|----|----|
| - | 0   | -1  | -2 | -3 | -4 | -5 | -6 | -7 |
| A | -1  | 1   |    |    |    |    |    |    |
| G | -2  | 0   |    |    |    |    |    |    |
| A | -3  | -1  |    |    |    |    |    |    |
| T | -4  | -2  |    |    |    |    |    |    |
| C | -5  | -3  |    |    |    |    |    |    |
| A | -6  | -4  |    |    |    |    |    |    |
| G | -7  | -5  |    |    |    |    |    |    |
| A | -8  | -6  |    |    |    |    |    |    |
| A | -9  | -7  |    |    |    |    |    |    |
| A | -10 | -8  |    |    |    |    |    |    |
| T | -11 | -9  |    |    |    |    |    |    |
| G | -12 | -10 |    |    |    |    |    |    |

$$M[i,j] = \max [ \begin{array}{l} M[i-1,j-1] + S(P[i],Q[j]), \\ M[i,j-1] + S(-, Q[j]) \\ M[i-1,j] + S(P[i], -) \end{array} ]$$

## 2. Fill Matrix

|   | –   | A   | T  | A  | G  | A  | A  | T  |
|---|-----|-----|----|----|----|----|----|----|
| – | 0   | -1  | -2 | -3 | -4 | -5 | -6 | -7 |
| A | -1  | 1   | 0  |    |    |    |    |    |
| G | -2  | 0   | 0  |    |    |    |    |    |
| A | -3  | -1  | -1 |    |    |    |    |    |
| T | -4  | -2  | 0  |    |    |    |    |    |
| C | -5  | -3  | -1 |    |    |    |    |    |
| A | -6  | -4  | -2 |    |    |    |    |    |
| G | -7  | -5  | -3 |    |    |    |    |    |
| A | -8  | -6  | -4 |    |    |    |    |    |
| A | -9  | -7  | -5 |    |    |    |    |    |
| A | -10 | -8  | -6 |    |    |    |    |    |
| T | -11 | -9  | -7 |    |    |    |    |    |
| G | -12 | -10 | -8 |    |    |    |    |    |

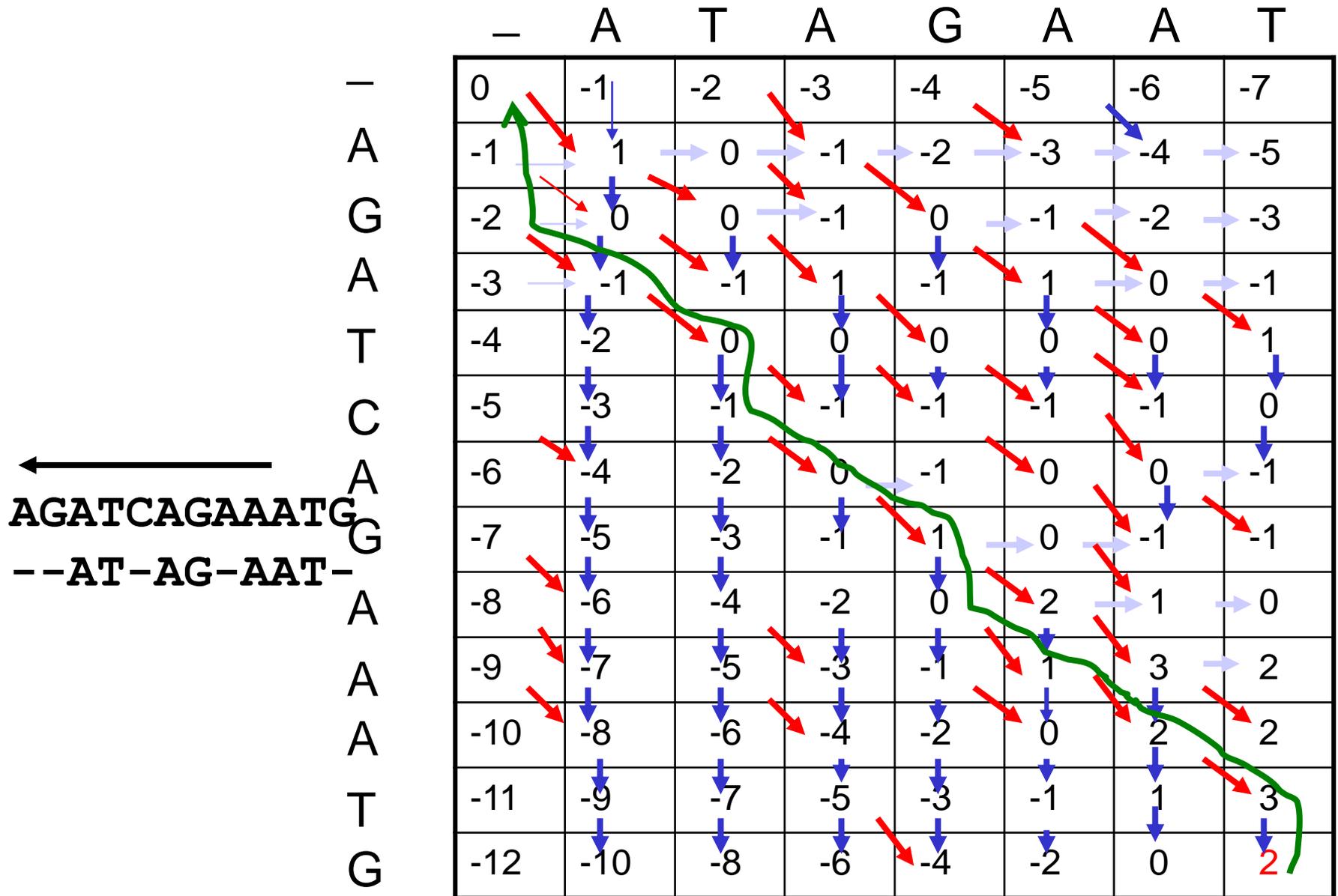
$$M[i,j] = \max [ \begin{array}{l} M[i-1,j-1] + S(P[i],Q[j]), \\ M[i,j-1] + S(-, Q[j]) \\ M[i-1,j] + S(P[i], -) \end{array} ]$$

## 2. Fill Matrix

|   | –   | A   | T  | A  | G  | A  | A  | T  |
|---|-----|-----|----|----|----|----|----|----|
| – | 0   | -1  | -2 | -3 | -4 | -5 | -6 | -7 |
| A | -1  | 1   | 0  | -1 | -2 | -3 | -4 | -5 |
| G | -2  | 0   | 0  | -1 | 0  | -1 | -2 | -3 |
| A | -3  | -1  | -1 | 1  | -1 | 1  | 0  | -1 |
| T | -4  | -2  | 0  | 0  | 0  | 0  | 0  | 1  |
| C | -5  | -3  | -1 | -1 | -1 | -1 | -1 | 0  |
| A | -6  | -4  | -2 | 0  | -1 | 0  | 0  | -1 |
| G | -7  | -5  | -3 | -1 | 1  | 0  | -1 | -1 |
| A | -8  | -6  | -4 | -2 | 0  | 2  | 1  | 0  |
| A | -9  | -7  | -5 | -3 | -1 | 1  | 3  | 2  |
| A | -10 | -8  | -6 | -4 | -2 | 0  | 2  | 2  |
| T | -11 | -9  | -7 | -5 | -3 | -1 | 1  | 3  |
| G | -12 | -10 | -8 | -6 | -4 | -2 | 0  | 2  |

$$M[i,j] = \max [ \begin{array}{l} M[i-1,j-1] + S(P[i],Q[j]), \\ M[i,j-1] + S(-, Q[j]), \\ M[i-1,j] + S(P[i], -) \end{array} ]$$

### 3. Trace Back



# Local vs. Global Alignment

- Global Alignment

```
--T--CC-C-AGT--TATGT-CAGGGGACACG-A-GCATGCAGA-GAC
|  || |  ||  |  |  |  |||  |  |  |  |  |  |  |  |  |
AATTGCCGCC-GTCGT-T-TTCAG-----CA-GTTATG-T-CAGAT--C
```

- Local Alignment—better alignment to find conserved segment

Transcription binding site

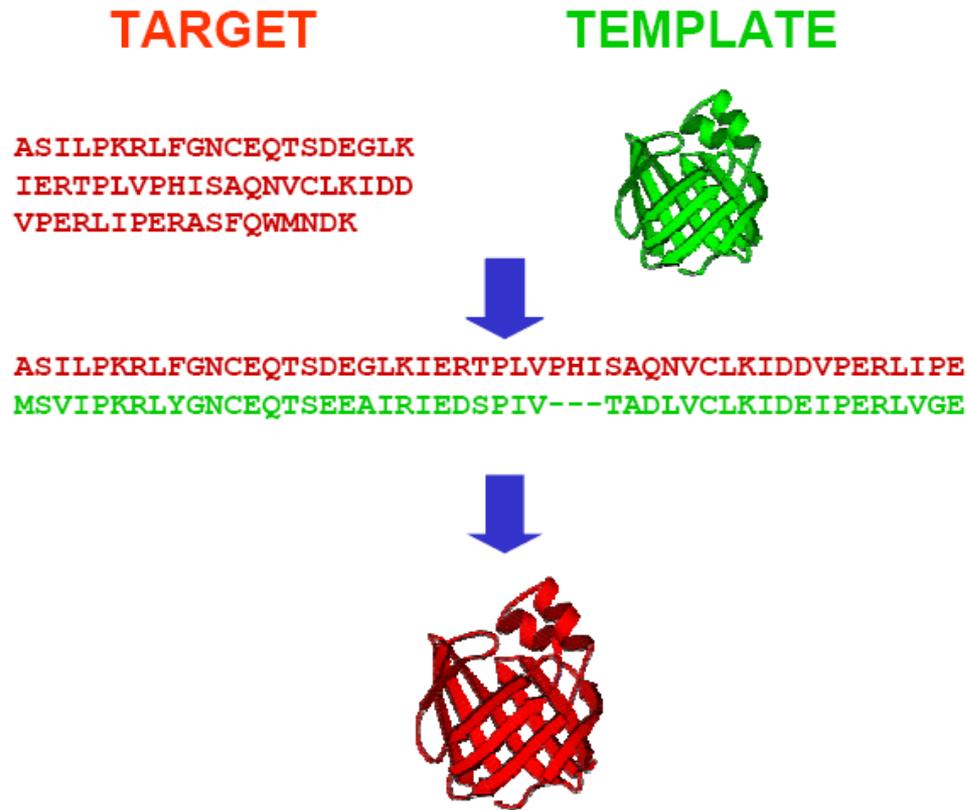
```
          ↗
tccCAGTTATGTCAGggggacacgagcatgcagagac
      ||| ||| ||| ||| |||
aattgccgccgtcgttttcagCAGTTATGTCAGatc
```

# Smith-Waterman Algorithm

Same dynamic program algorithm as global alignment except for three differences.

1. All negative scores is converted to 0
2. Alignment can start from anywhere in the matrix
3. Alignment can end at anywhere in the matrix

# Application Example (Alignment – Structure)



Source: A. Fisher, 2005

# Global and Local Alignment Tools

- NEEDLE (global alignment)

<http://bioweb.pasteur.fr/seqanal/interfaces/needle.html>

- WATER (local alignment)

<http://bioweb.pasteur.fr/seqanal/interfaces/water.html>

# Scoring Matrix

- How to accurately measure the similarity between amino acids (or nucleotides) is one key issue of sequence alignment.
- For nucleotides, a simple identical / not identical scheme is mostly ok.
- Due to various properties of amino acids, it is hard and also critical to measure the similarity between amino acids.

# Evolutionary Substitution Approach

- During evolution, the substitution of similar (or dissimilar) amino acids is more (or less) likely to be selected within protein families than random substitutions (M. Dayhoff)
- The frequency / probability one residue substitutes another one is an indicator of their similarity.

# PAM Scoring Matrices

(M. Dayhoff)

- Select a number of protein families.
- Align sequences in each family and count the frequency of amino acid substitution of each column. The frequency is used to compute the empirical substitution probability of which residue  $i$  substitutes residue  $j$  ( $P_{ij}$ ).
- Similarity score is ratio of observed substitution probability over the random substitution probability.  
 $S(i,j) = \log(P_{ij} / (P_i * P_j))$ .  $P_i$  is the observed probability of residue  $i$  and  $P_j$  is the observed probability of residue  $j$
- PAM: Point Accepted Mutation

# A Simplified Example

ACGTCGAGT  
 ACCACGTGT  
 CACACTACT  
 ACCGCATGA  
 ACCCTATCT  
 TCCGTAACA  
 ACCATAAGT  
 AGCATAAGT  
 ACTATAAGT  
 ACGATAAGT

| Chars | Prob.  |
|-------|--------|
| A     | 6 / 10 |
| C     | 1 / 10 |
| G     | 2 / 10 |
| T     | 1 / 10 |

Substitution Frequency Table

|   | A  | C | G  | T |
|---|----|---|----|---|
| A | 30 | 6 | 12 | 6 |
| C | 6  | 0 | 2  | 1 |
| G | 12 | 2 | 1  | 2 |
| T | 6  | 1 | 2  | 0 |

Total number of substitutions: 90

|   | A   | C   | G   | T   |
|---|-----|-----|-----|-----|
| A | .33 | .07 | .14 | .07 |
| C | .07 | 0   | .02 | .01 |
| G | .14 | .02 | .01 | .02 |
| T | .07 | .01 | .02 | 0   |

$$P(A \leftrightarrow C) = 0.07 + 0.07 = 0.14$$

# A Simplified Example

ACGTCGAGT  
 ACCACGTGT  
 CACACTACT  
 ACCGCATGA  
 ACCCTATCT  
 TCCGTAACA  
 ACCATAAGT  
 AGCATAAGT  
 ACTATAAGT  
 ACGATAAGT

| Chars | Prob.  |
|-------|--------|
| A     | 6 / 10 |
| C     | 1 / 10 |
| G     | 2 / 10 |
| T     | 1 / 10 |

Substitution Frequency Table

|   | A  | C | G  | T |
|---|----|---|----|---|
| A | 30 | 6 | 12 | 6 |
| C | 6  | 0 | 2  | 1 |
| G | 12 | 2 | 1  | 2 |
| T | 6  | 1 | 2  | 0 |

Total number of substitutions: 90

|   | A   | C   | G   | T   |
|---|-----|-----|-----|-----|
| A | .33 | .07 | .14 | .07 |
| C | .07 | 0   | .02 | .01 |
| G | .14 | .02 | .01 | .02 |
| T | .07 | .01 | .02 | 0   |

$$P(A \leftrightarrow C) = 0.07 + 0.07 = 0.14$$

$$S(A, C) = \log(0.14 / (0.6 * 0.1)) = 0.36$$



# BLOSUM Matrices

(Henikoff and Henikoff)

- PAM matrices don't work well for aligning evolutionarily divergent sequences.
- BLOSUM: BLOcks SUBstitution Matrix
- PAM based on observed mutations throughout global alignment. BLOSUM based on highly conserved local regions /blocks without gaps.
- BLOSUM<sub>n</sub> is a matrix calculated from proteins share at most n% identity. BLOSUM62 is the most widely used matrix (BLAST, PSI-BLAST, CLUSTALW)

```

-VLSPADKTNVKAANGKVGAAHAGEYGAEALERMFLSFPTTKTYFPHF-DLS-----HGSA
-VLSAADKTNVKAAWSKVGGHAGEYGAEALERMFLGFPTTKTYFPHF-DLS-----HGSA
VHLTPEEKSAVTALWGKVN--VDEVGGEALGRLLVYPWTQRFFESFGDLSTPDAVMGNP
VQLSGEEKAAVLALWDKVN--EEEVGGEALGRLLVYPWTQRFFDSFGDLSPGAVMGNP
-VLSEGEWQLVLHVWAKVEADVAGHGQDILIRLFKSHPETLEKFDRFKHLKTEAEMKASE

```

Block 1

Block2

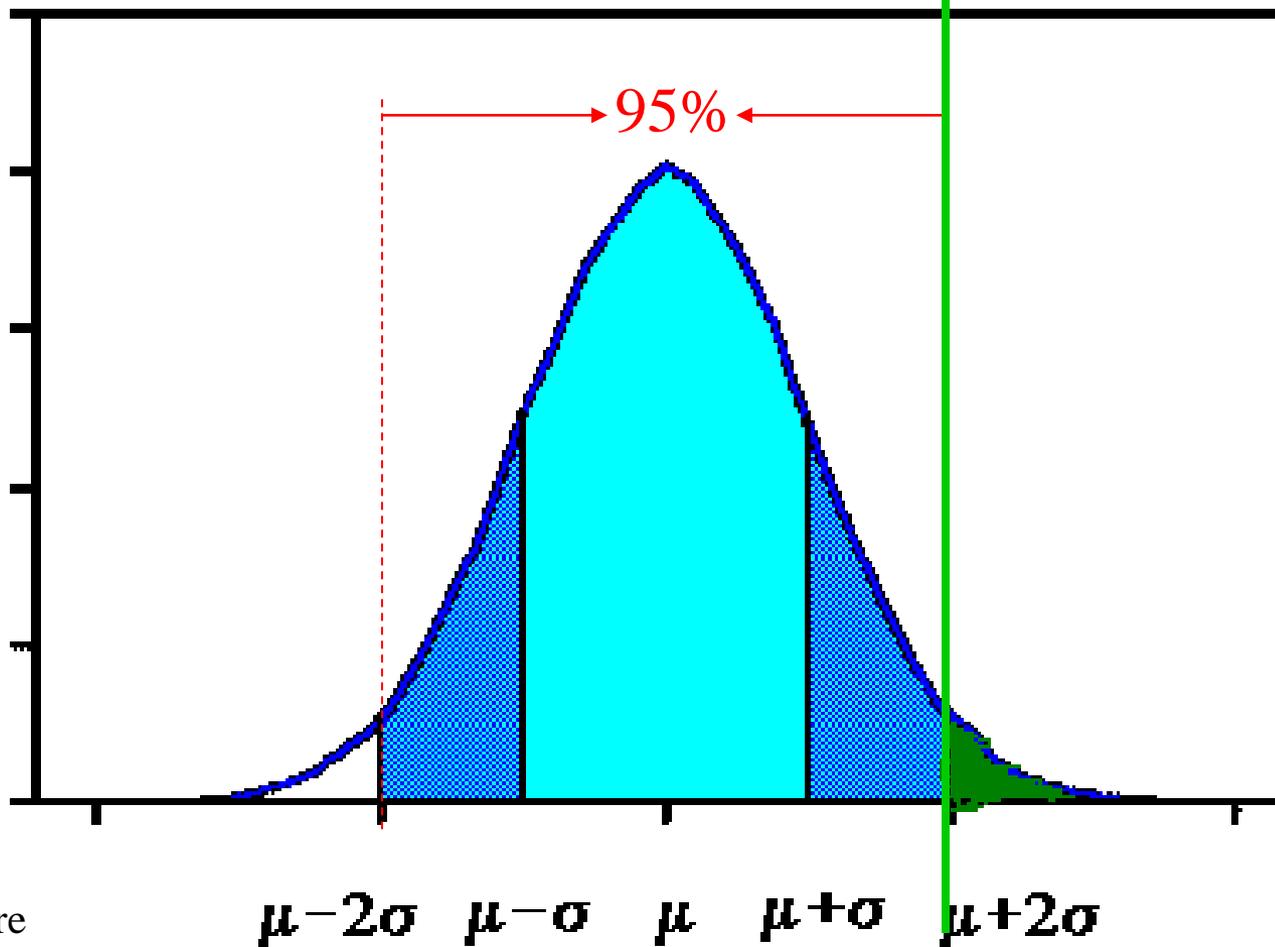


# Significance of Sequence Alignment

- Why do we need significant test?
- Mathematical view: unusual versus “by chance”
- Biological view: evolutionary related or not?

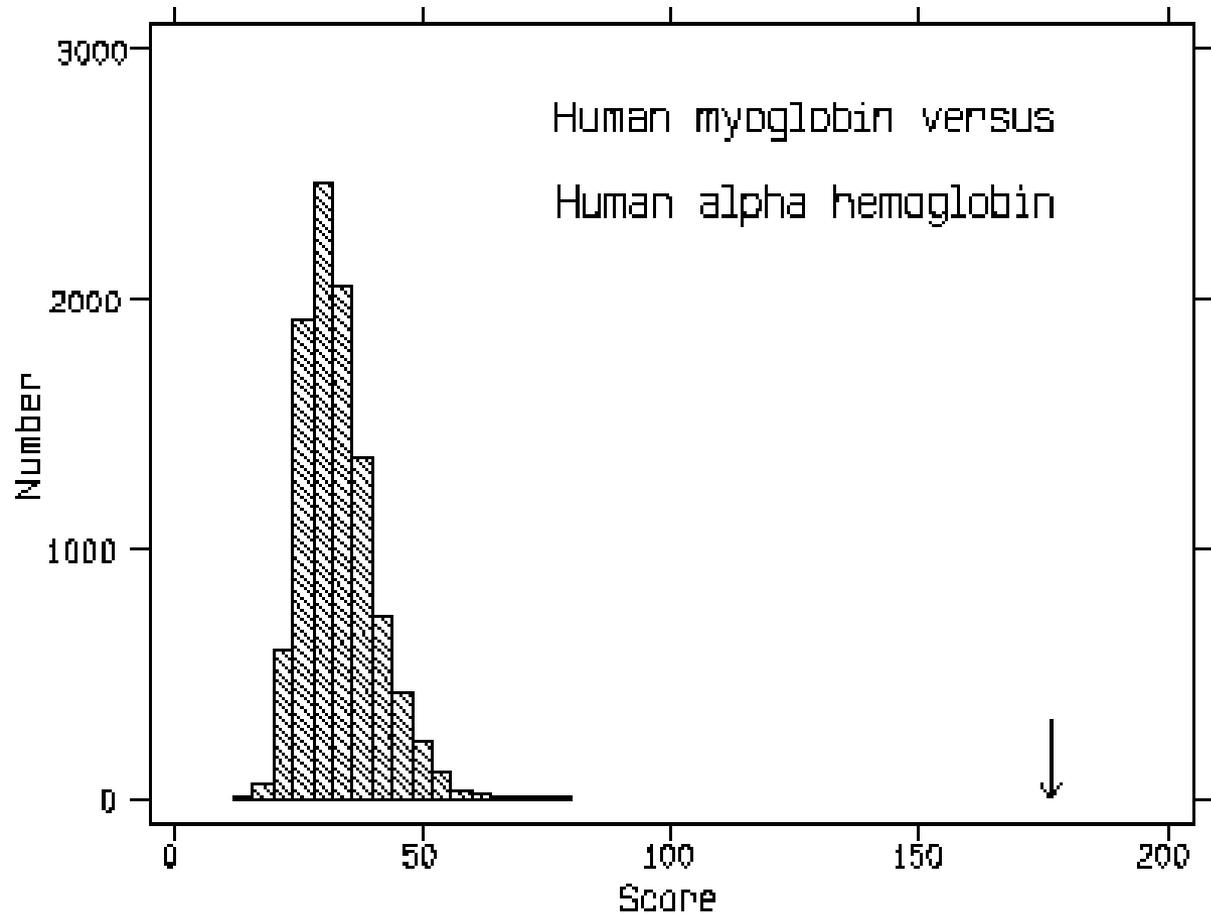
# Randomization Approach

- Randomization is a fundamental idea due to Fisher.
- Randomly permute chars within sequence P and Q to generate new sequences (P' and Q'). Align new sequences and record alignment scores.
- Assuming these scores obey normal distribution, compute mean ( $\mu$ ) and standard deviation ( $\sigma$ ) of alignment scores



## Normal distribution of alignment scores of two sequences

- If  $S = u + 2\sigma$ , the probability of observing the alignment score equal to or more extreme than this by chance is 2.5%, e.g.,  $P(S \geq u + 2\sigma) = 2.5\%$ . Thus we are 97.5% confident that the alignment score is significant (not by chance).
- For any score  $x$ , we can compute  $P(S \geq x)$ , which is called p-value.



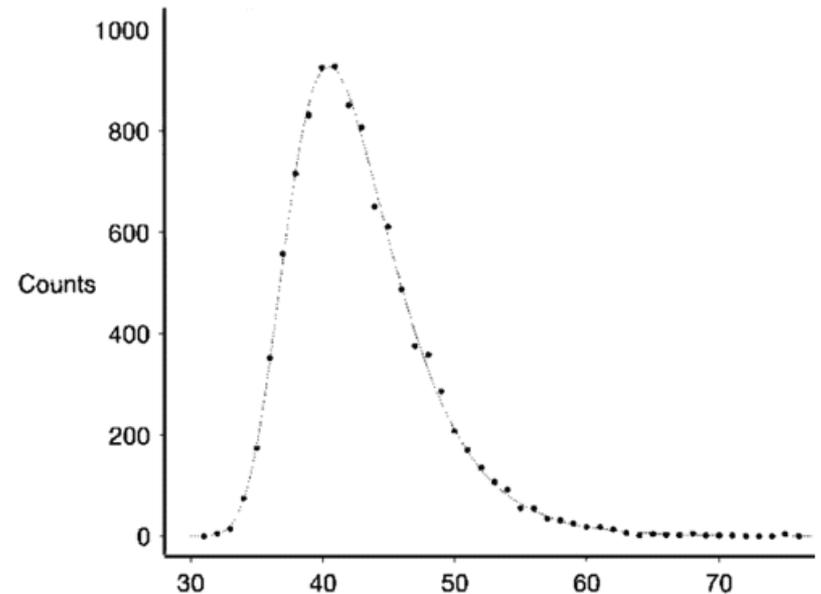
**Figure:** Histogram of alignment scores

# Model-Based Approach (Karlin and Altschul)

<http://www.people.virginia.edu/~wrp/cshl02/Altschul/Altschul-3.html>

- Extreme Value  
Distribution

$$P(S \geq x) = 1 - \exp(-Kmn e^{-\lambda x})$$



K and lamda are statistical parameters depending on substitution matrix. For BLOSUM62, lamda=0.252, K=0.35

# P-Value

- $P(S \geq x)$  is called **p-value**. It is the probability that random sequences has alignment score equal to or bigger than  $x$ .
- Smaller  $\rightarrow$  more significant.

# Problems of Using Dynamic Programming to Search Large Sequence Database

- Search homologs in DNA and protein database is often the first step of a bioinformatics study.
- DP is too slow for large sequence database search such as Genbank and UniProt. Each DP search can take hours.
- Most DP search time is wasted on unrelated sequences or dissimilar regions.
- Developing fast, practical sequence comparison methods for database search is important.

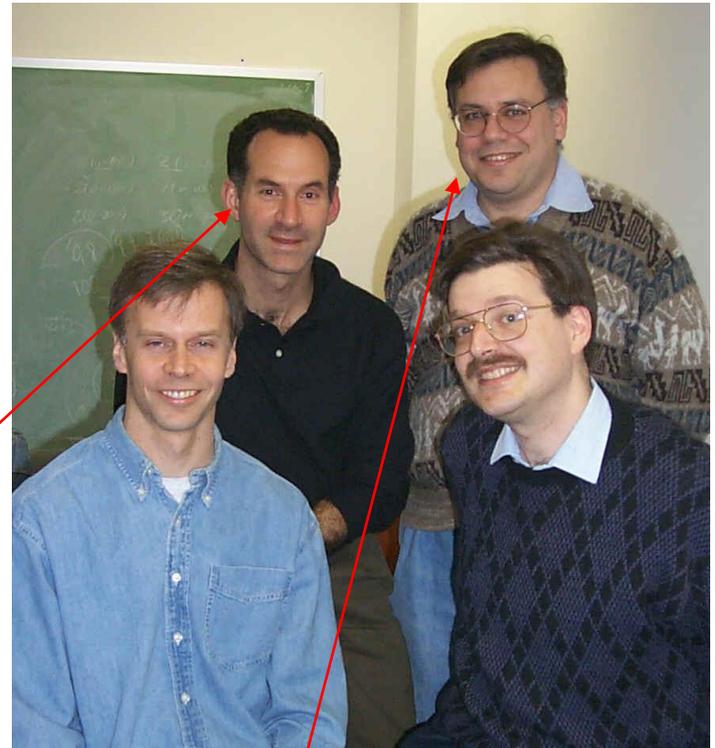
# Fast Sequence Search Methods

- All successful, rapid sequence comparison methods are based on a simple fact: similar sequences /regions **share some common words**.
- First such method is FASTP (Pearson & Lipman, 1985)
- Most widely used methods are BLAST (Altschul et al., 1990) and PSI-BLAST (Altschul et al., 1997).

# Basic Local Alignment Search Tool

(S. Altschul, W. Gish, W. Miller, E. Meyer and D. Lipman)

1. Compile a list of words for a query
2. Scan sequences in database for word hits
3. Extending hits



David Lipman

Stephen Altschul

# Compile Word List

- Words: w-mer with length w.
- Protein 4-mer and DNA 12-mer

**Query:**

DSRSKGEPRDSGTLQSQEAKAVKKTSLFE

**Words:** DSRS, SRSK, RSKG, KGEP....

# Example of extension

Query: **DSRSK**GEPRDSGTLQSQEAKAVKKTSLFE

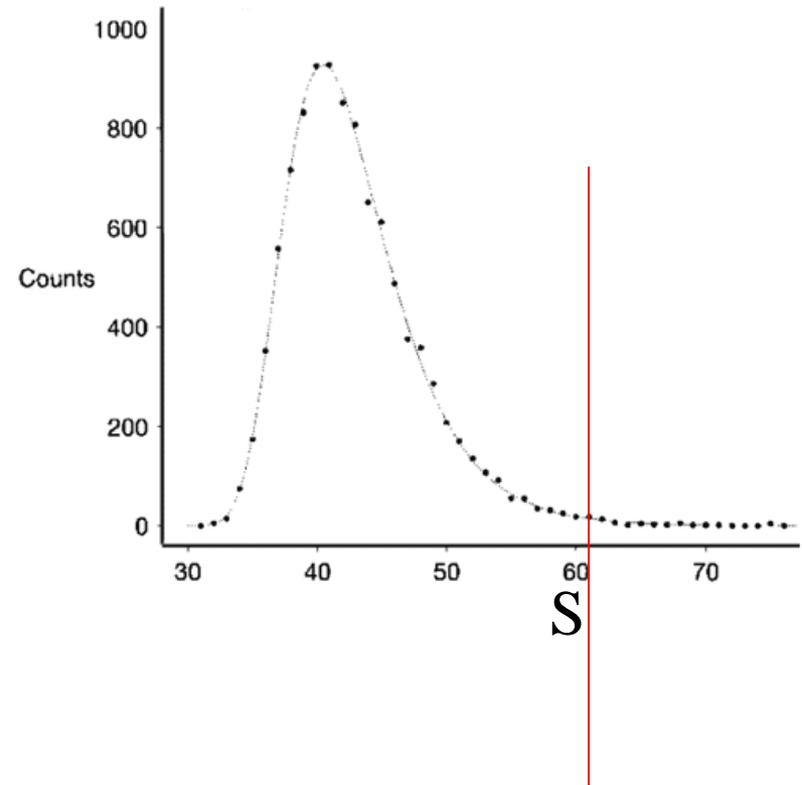
Words: DSRS, **SRSK**, RSKG, KGEP....

Database Sequence: P**ESRSK**GEPRDS**G**KKQMDSOKPD  


Maximum Segment Pair: **ESRSKGEPRDSG**

# P-Value and E-Value

- P-value
- E-value = database size \* p-value
- Common threshold: 0.01



$$\text{P-value} = \text{Prob}(\text{score} \geq S)$$

# Usage of BLAST

- Versions: BLASTP, BLASTN, BLASTX (translated)
- Sequence Databases: NR, PDB, SwissProt, Gene databases of organisms, or your own databases
- Expectation value
- Low complexity
- Similarity matrix (PAM or BLOSUM)
- Output format

# NCBI Online Blast

The image shows a screenshot of the NCBI BLAST website. At the top, there is a browser address bar with the URL <http://www.ncbi.nlm.nih.gov/BLAST/>. Below the address bar is a search bar with the text "ncbi" and a search button. The main content area is titled "NCBI → BLAST" and includes a navigation menu on the left with sections: "About", "More info", "Software", and "Other resources". The main content area is divided into four quadrants: "Nucleotide", "Protein", "Translated", and "Genomes", each with a list of search options.

Latest news: 7 May 2006 : BLAST 2.2.14 released

**About**

- Getting started
- News
- FAQs

**More info**

- NAR 2004
- NCBI Handbook
- The Statistics of Sequence Similarity Scores

**Software**

- Downloads
- Developer info

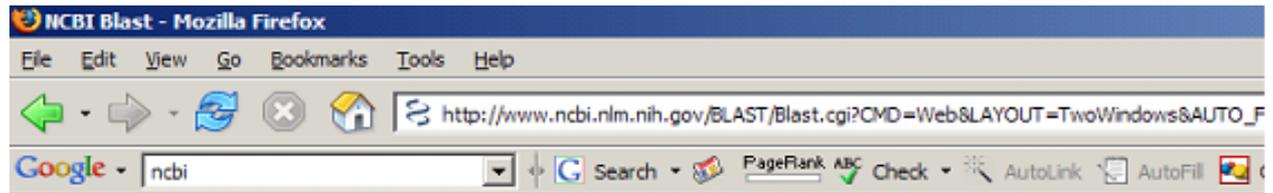
**Other resources**

- References
- NCBI Contributors
- Mailing list
- Contact us

**The Basic Local Alignment Search Tool (BLAST)** finds regions of local similarity between sequences. The program compares nucleotide or protein sequences to sequence databases and calculates the statistical significance of matches. BLAST can be used to infer functional and evolutionary relationships between sequences as well as help identify members of gene families.

|  |  |
|--|--|
| <b>Nucleotide</b> <ul style="list-style-type: none"><li>• Quickly search for highly similar sequences (megablast)</li><li>• Quickly search for divergent sequences (discontiguous megablast)</li><li>• Nucleotide-nucleotide BLAST (blastn)</li><li>• Search for short, nearly exact matches</li><li>• Search trace archives with megablast or discontiguous megablast</li></ul> | <b>Protein</b> <ul style="list-style-type: none"><li>• Protein-protein BLAST (blastp)</li><li>• Position-specific iterated and pattern-hit initiated BLAST (PSI- and PHI-BLAST)</li><li>• Search for short, nearly exact matches</li><li>• Search the conserved domain database (rpsblast)</li><li>• Protein homology by domain architecture (cdart)</li></ul> |
| <b>Translated</b> <ul style="list-style-type: none"><li>• Translated query vs. protein database (blastx)</li><li>• Protein query vs. translated database (tblastn)</li><li>• Translated query vs. translated database (tblastx)</li></ul>  | <b>Genomes</b> <ul style="list-style-type: none"><li>• Human, mouse, rat, chimp, cow, pig, dog, sheep, cat</li><li>• Chicken, puffer fish, zebrafish</li><li>• Fly, honey bee, other insects</li><li>• Microbes, environmental samples</li><li>• Plants, nematodes</li><li>• Fungi, protozoa, other eukaryotes</li></ul>                                       |

# DNA Blast



[Search](#)

[Set subsequence](#) From:  To:

[Choose database](#)

Now: **BLAST!** or **Reset query** **Reset all**

**Options** for advanced blasting

[Limit by entrez query:](#)  or select from:

[Choose filter](#)  Low complexity  Human repeats  Mask for lookup table only  Mask lower case

[Expect](#)

[Word Size](#)

[Other advanced](#)

# Protein Blast

[Search](#)

[Set subsequence](#) From:  To:

[Choose database](#)

[Do CD-Search](#)

Now: **BLAST!** or

## Options for advanced blasting

[Limit by entrez query](#)  or select from:

[Compositional adjustments](#)

[Choose filter](#)  Low complexity  Mask for lookup table only  Mask lower case

[Expect](#)

[Word Size](#)

MASKTIKIMPVGDSCTEGMGGGEMGSYRTELYRLLTQAGLSIDFVGSQRSGPSSLPDKDH  
 EGHSGWTIPQIASNINNLNTHNPDVVFVFLWIGNDLLLNGNLNATGLSNLIDQIFTVKEN  
 VTLFVADYYPWPEAIKQYNAVIGIVQQKANAGKKVYFVKLSEIQFDRNTDISWDGLHLS  
 EIGYKKIANIWIYKTIDILRALAGE

[Search](#)

[Set subsequence](#) From:  To:

[Choose database](#)

[Do CD-Search](#)

Now: **BLAST!** or **Reset query** **Reset all**

The request ID is

**Format!** or **Reset all**

| Sequences producing significant alignments:                                      | Score (Bits)         | E Value |
|--|----------------------|---------|
| <a href="#">gi 67876011 ref ZP_00505069.1 </a> Lipolytic enzyme, G-D-S-L:Clos... | <a href="#">344</a>  | 1e-93   |
| <a href="#">gi 121831 sp P15329 GUNX_CLOTM</a> Putative endoglucanase X (EGX)... | <a href="#">227</a>  | 2e-58   |
| <a href="#">gi 35213333 dbj BAC90705.1 </a> gll2764 [Gloeobacter violaceus PC... | <a href="#">103</a>  | 5e-21   |
| <a href="#">gi 89241797 emb CAJ81036.1 </a> putative xylanase [Actinoplanes sp.  | <a href="#">90.9</a> | 3e-17   |
| <a href="#">gi 46123721 ref XP_386414.1 </a> hypothetical protein FG06238.1 [... | <a href="#">87.4</a> | 3e-16   |
| <a href="#">gi 111057360 gb EAT78480.1 </a> hypothetical protein SNOG_14243 [Pha | <a href="#">83.2</a> | 7e-15   |
| <a href="#">gi 90294376 ref ZP_01213970.1 </a> hypothetical protein Bpse17_02... | <a href="#">82.0</a> | 1e-14   |
| <a href="#">gi 52209736 emb CAH35705.1 </a> putative exported oxidase [Burkho... | <a href="#">81.3</a> | 2e-14   |
| <a href="#">gi 76579113 gb ABA48588.1 </a> galactose oxidase-like protein [Bu... | <a href="#">81.3</a> | 3e-14   |
| <a href="#">gi 111225445 ref YP_716239.1 </a> putative Glycosyl hydrolase [Fr... | <a href="#">79.3</a> | 9e-14   |

## Matched sequences ranked by score and evalue

# Output Format

```
>  gi|35213333|dbj|BAC90705.1|  gll2764 [Gloeobacter violaceus PCC 7421]
gi|37522333|ref|NP\_925710.1|  hypothetical protein gll2764 [Gloeobacter violaceus PCC 7421]
Length=559

Score = 103 bits (256), Expect = 5e-21, Method: Composition-based stats.
Identities = 89/194 (45%), Positives = 115/194 (59%), Gaps = 12/194 (6%)

Query 7 KIMPVGDSCTEGMGGGEMGSYRTELYRLLTQAGLSIDFVGSQRSGPSSLPDKDHGEGHSGW 66
      K+MP+GDS TEG G YRT+L+ L G +DFVGSQ SGPSSL DK+HEGH G+
Sbjct 108 KVMPLGDSITEGFTVS--GGYRTDLWNSLVSEGSNADFVGSQSSGPSSLSDKNHGHPGY 165

Query 67 TIPQIASNINNLNTHNPDVVFVFLWIGNDLLLNGN--lnatglsnlIDQIFTVKPNVTLF 124
      I QIA I++WL + P+ V L IG ND+ N + LS LIDQIF ++ +V L+
Sbjct 166 FIDQIADGIDDWLPKYKPEVLLLLIGTNDIEKNNDPGGAPGRLSALIDQIFALRSSVKLY 225

Query 125 VADYYPWPE-AIKQ----YNAVIGIVQQKANAGKKVYFVKLSEIQFDRNTDISWDGLHL 179
      VA P + AI Q YNA IPGIV K GKKV +V + D++ D +H
Sbjct 226 VASIPPADDSAINQRVLDYNAIIPGIVNGKITQGKKVYVVDIYNAL--TTADLA-DIVHP 282

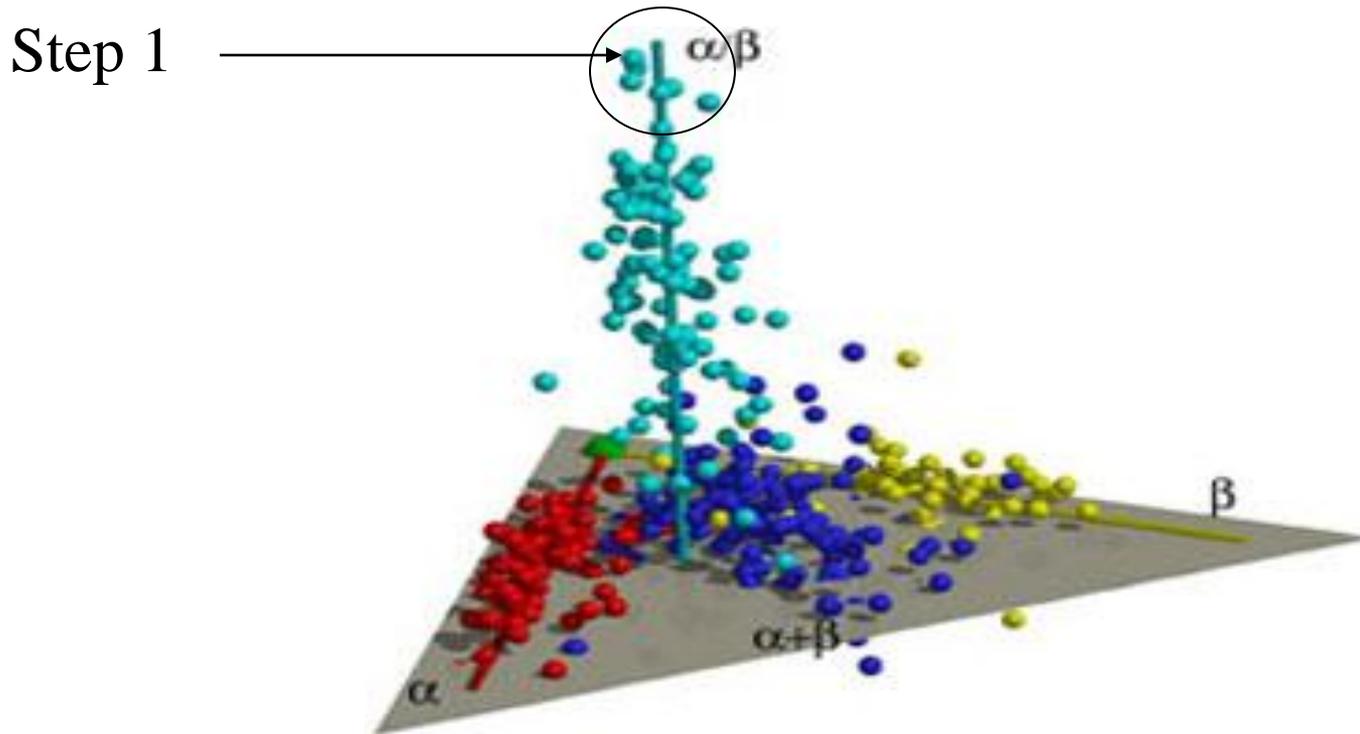
Query 180 SEIGYKKIANIWIYK 193
      GY KIA+ W++
Sbjct 283 DAEGYAKIADRWFE 296
```

## Significant local alignments

# Database Search Using Sequence Profiles

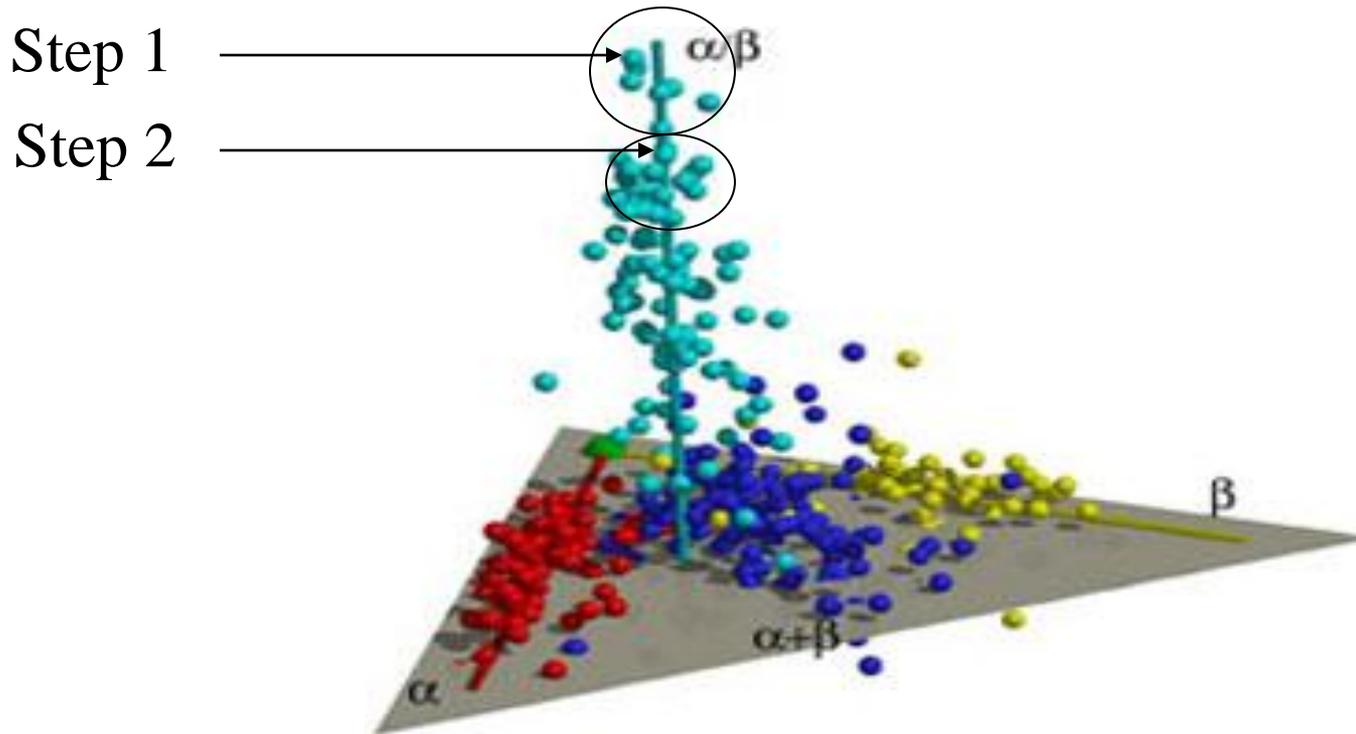
- Multiple related sequences in protein family and super family (profile)
- More data, more robust, more sensitive
- Consider a group of related sequences (profile) is a **POWERFUL** idea

# Why does a family of sequences help?



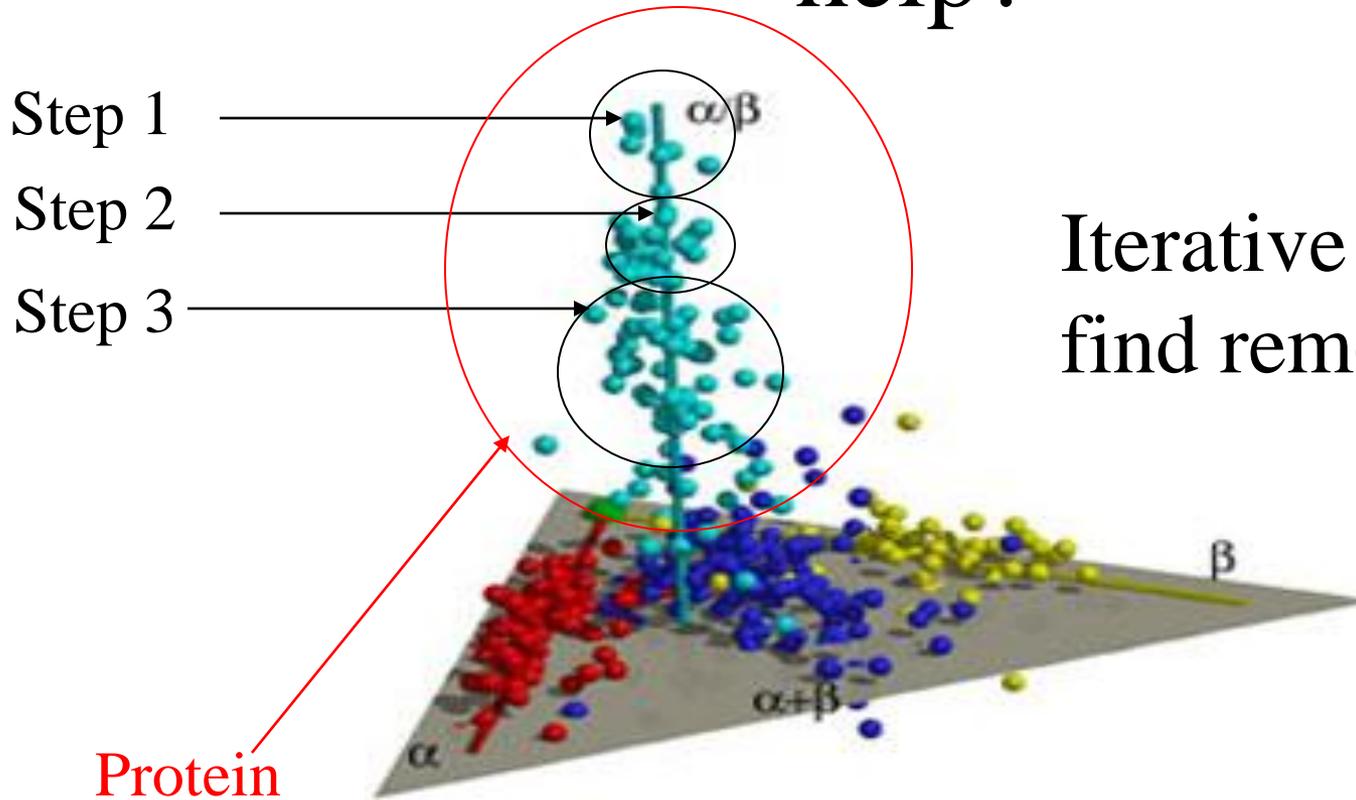
**Protein Universe**

# Why does a family of sequences help?



**Protein Universe**

# Why does a family of sequences help?



Step 1

Step 2

Step 3

Iterative search helps find remote homologs.

Protein Family

Protein Universe

# PSI-BLAST Algorithm

- Use BLAST to search database. Use significantly matched sequences to construct a profile / PSSM
- Repeat
  - Use PSSM to search database
  - Use significant matched sequences to construct a PSSM
- Until no new sequence is found or reach the maximum number of iterations.

# Use PSI-BLAST Software

- Download: <http://130.14.29.110/BLAST/download.shtml>
- Command:

```
blastpgp -i seq_file -j iteration -h include_evalue_threshold -e report_evalue_threshold -d database -o output_file
```

**-i:** input sequence file in FASTA format

**-j:** number of iterations

**-d:** sequence database

**-h:** cut-off e-value of including a sequence into PSSM (profile)

**-e:** cut-off e-value of reporting a sequence

**-o:** output file